Total NMR Assignment of Furanditerpene Derivatives from *Pterodon Polygalaeflorus Benth.*

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Abstract: Pterodon polygalaeflorus Benth, popularly known as "Sucupira-branca", is a well-known tree in the central region of Brazil. The seeds of this tree are used in folk medicine as anti-rheumatic, antiinflammatory and analgesic preparations. From the hexane extract of its fruits, 6α.7βdihydroxyvouacapan-17 β -oic acid (ADV), a furanditerpene that presents anti-inflammatory, analgesic, and plant growth regulatory activities was isolated. ADV (1) and some derivatives have previously been studied to obtain some clues about chemical structure-biological activity relationship. Some ADV esters containing an OH group at C-6 have already been obtained. Both hydroxyl and carbonyl groups can receive hydrogen bonds, while the hydroxyl group can also donate a hydrogen bond. With the aim of mapping the biological receptor, the replacement of the OH group at C-6 of ADV by a carbonyl group was made. Therefore, the 6-oxo-voucapan- 7α , 17β -lactone (3) derivative was prepared. Here we described the NMR studies of four ester derivatives obtained from (3), which were as follows: methyl 6- $0 \times 0.7 \alpha$ -hydroxyvouacapan-17 β -oate (4), ethyl 6-0x0-7 β -hydroxyvouacapan-17 β -oate (5), propyl 6-0x0- 7α -hydroxyvouacapan-17 β -oate (**6**), 2-methoxy-ethyl 6-oxo- 7α -hydroxyvouacapan-17 β -oate (**7**). Their resonances were unequivocally assigned by the use of 1D (¹H and ¹³C NMR, DEPT-135, NOE difference), and 2D (COSY, HMQC, HMBC) NMR techniques.

The seeds of *Pterodon polygalaeflorus* Benth, popularly known as "*Sucupira-branca*" are used in folk medicine as anti-rheumatic, anti-inflammatory and analgesic preparations.¹ From the hexane extract of its fruits it was isolated the 6α , 7β -dihydroxyvouacapan- 17β oic acid (ADV, **1**, Figure 1)², a furanditerpene that presents anti-inflammatory, analgesic, and plant growth regulatory activities.^{2,3}

To obtain some information with respect to the chemical structure-biological activity relationship, ADV, as well as its lactone (6α hydroxyvoucapan- 7β , 17β -lactone, HVL, **2**, Figure 1) and some of their ester and amide derivatives were previously studied.⁴ The NH, OH, and CO groups are able to receive hydrogen bonds, while the NH and OH groups can also donate a hydrogen to hydrogen bonds.

To map the receptor structure, the replacement of the OH group at C-6 of the ADV lactone derivative, HVL (2) by a carbonyl group was undertaken. The total NMR characterization of four esters (4-7) derived from the POL (3, Figure 1) is described herein.

The total NMR assignment of the esters **4-7** is discussed below. The synthesis of these compounds has already been reported.⁵ The 1D and 2D NMR techniques of ¹H, ¹³C, DEPT-135, NOE difference; COSY, HMQC, and HMBC were used.

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¹H and ¹³C NMR spectra were recorded on a Bruker *AVANCE* DRX 400 spectrometer, in CDCl₃. Chemical shifts are reported in parts per million (δ units), relative to TMS as internal standard, and scalar coupling constants (*J*) are reported in hertz. Pulse conditions were as follows: for ¹H NMR spectra - dwell time (DW) 149.600 µs, acquisition time (AQ) 3.985 s, number of transients (NS) 16, recycle delay (RD) 1.000 s; for ¹³C NMR spectra – DW 31.400 µs, AQ 2.058 s, NS 1024, RD 2.000 s, decoupling multiple resonance method Waltz-16; for COSYGR – DW 125.000 µs, AQ 0.116 s, NS 1, DS 8, RD 2.000 s, data points (TD) 1024 (F2) and 256 (F1); HMQC – DW 54.400

μs, AQ 0.058, NS 16, RD 2.000 s, TD 1024 (F2) and 512 (F1); HMBC – DW 54.400 μs, AQ 0.223 s, NS 32, RD 2.000, delay for long range coupling (D6) 0.070 s, TD 2048 (F2) and 512 (F1).

Figure 1 shows the synthetic route to obtain the esters **4-7**. The synthesis accomplishment was verified by the appearance of the hydroxyl group hydrogen resonance around δ 3.6 in the ¹H NMR spectra of all the esters. Other pieces of evidence for the obtention of the esters are provided by the ¹H and ¹³C resonances due to the R groups shown in Figure 1 from **4** to **7**.



Figure 1. Synthetic route of C-6 ceto ester derivatives (4-7) of ADV.^{6,7}

The HMQC contour map was useful to attribute resonance signals to nonmagnetically equivalent *gem* hydrogens, since they resonate at different chemical shifts. To illustrate, an expansion of ester **6** ¹³C NMR spectrum is shown in Figure 2, where C-11 and C-22 present the same δ_C 22.0.



Figure 2. Partial ¹³C NMR spectrum of ester 6 in CDCI₃ at 100 MHz.

Since both signals are due to CH₂, they were also overlapped in DEPT-135. Therefore, they were assigned from the HMQC analysis (upper right side of Figure 2) by the correlations of $\delta_{\rm C}$ 22.0 with H-11ax (δ 2.50), H-11eq (δ 2.75), and H-22 (δ 1.70). The HMBC contour maps were useful for the assignment of non-hydrogenated carbon signals and also for distinguishing the CH₃-20 ($\delta_{\rm H}$ 0.91; $\delta_{\rm C}$ 15.1) from both CH₃-18 ($\delta_{\rm H}$ 0.97; $\delta_{\rm C}$ 32.5) and CH₃-19 ($\delta_{\rm H}$ 1.32; $\delta_{\rm C}$ 22.2) signals. Thus, H-20 correlates via ${}^{3}J$ with both C-1 (δ_{C} 38.6) and C-9 ($\delta_{\rm C}$ 48.1) signals, and via ²J with C-10 (δ 42.8). The NOE difference experiment distinguished between CH₃-18 and CH₃-19 signals. NMR techniques confirmed the synthesis and the structures of the esters.

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